

Characterization of the Leukaemia-Associated ETO Homologous

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Abstract: MTG16, MTGR1 and ETO are nuclear transcriptional corepressors of the human ETO protein gang. MTG16 is ensnared in hematopoietic improvement and in controlling megakaryopoiesis. Moreover, ETO homologue qualities are 3'participants in leukemia combinations created by chromosomal translocations mindful of hematopoietic dysregulation. The proposed cooperations of the ETO homologues may have suggestions for the onset of leukemia, since it opens up for an AML1-ETO interceded aggravation of ETO homologue capacity and a regulation of AML1-ETO capacity by the ETO homologues. In this review we tried to identify the outcomes of several reviews that are discussing Characterisation of the leukaemia-associated ETO homologous.

Keywords: Leukaemia-Associated ETO Homologous, AML1-ETO.

1. INTRODUCTION

Haematopoiesis is a strictly controlled procedure, prompting the development of cells important for a utilitarian safe resistance, oxygen transport and hemostasis. The high-rate generation of blood cells and the firm control of the distinctive procedures roll out the framework helpless against improvements that influence the adjusted harmony between proliferation, differentiation and cell death (Levanon D et al, 1989).

Neoplasias of the blood shaping organs are delegated acute and chronic leukemias that are further separated into myeloid and lymphoid classifications. Regular for acute myeloid leukemia is that the ordinary generation of blood cells is disturbed by a clonal extension of harmful cells that ordinarily are captured in differentiation and are pretty much impervious to apoptosis. These outcomes in an amassing of youthful blastlike cells. Interestingly, chronic leukemia cells, for instance in CML, show nonstop hematopoietic differentiation. The patients with CML might have couple of manifestations at an early stage, though acute leukemia is a quickly advancing illness and will bring about casualty if left untreated (Gahrton, G et al, 1999). In strong tumors varieties of heterogeneous hereditary distortions have been accounted for, comprising of chromosomal erasures and translocations, quality intensifications and point transformations (Mitelman et al, 2006). In leukemia, a procured chromosomal translocation is a run of the mill hereditary distortion (Rowley et al, 1990). More than 300 chromosomal translocations have been portrayed in leukemia, however many less leukemic phenotypes have been characterized. This shows distinctive chromosomal translocations might have comparable phenotypes maybe reflecting comparative association of cellular flagging pathways for proliferation, differentiation and practicality (Kelly et al, 2002). A chromosomal translocation can bring about lost capacity from haploinsufficiency or an addition of capacity through a chimeric protein with novel properties.

In leukemia, the creation of full grown blood cells is disturbed at some level, prompting an aggregation of nonmature blood cells to the detriment of adult, practical cells. Every now and again, leukemia is connected with a chromosomal translocation including a haematopoietic key translation variable. The translocation results in the statement of a distorted combination protein with various properties and capacities than those of the ordinary interpretation component. t (8;21) is the most continuous translocation found in patients with acute myeloid leukemia (Miyoshi H, et al, 1993). It results in the declaration of the chimaeric protein Acute Myeloid Leukemia 1 (AML1)- ETO. AML1 capacities as a vital translation component in the haematopoietic system. The normal function of ETO, on the other hand, is largely unknown. The ETO component of AML1-ETO possesses the fusion protein with features that are crucial for its actions. The focus of this

work is on the function of ETO and its two homologues, Myeloid Translocation Gene Related protein 1 (MTGR1) and Myeloid Translocation Gene 16 (MTG16), with the aim to elucidate more about their functions in ordinary life as well as in leukaemogenesis.

Objectives:

AML1-ETO requires the ETO moiety to be able to interfere with transcriptional regulation. Summary of the cell biology of the AML1-ETO discussed in the previous section, it emphasizes changes of gene regulation in hematopoietic stem cells leading to a partial differentiation block. It also emphasizes the AML1-ETO-mediated growth arrest that has to be overcome by “second hits” for leukemia to occur. In addition, this study aim is to detect the Characterisation of the different types of leukaemia-associated ETO homologous, by reviewing the previous studies that were discussing this topic.

2. METHODOLOGY

This study is a review of literature, we have performed a comprehensive search that was undertaken by searching through the US National Library of Medicine (Pubmed), The following criteria had to be met for the publication to be selected topic and all these studies which were discussing the **Characterisation of the leukaemia-associated ETO homologous** was included most important studies that were conducted up to December 2015, our search ETO homologous, Leukaemia, AML, MLL.

Then we finally analysis the data and results of each included study to come out with the main and useful summarized results about the **Characterisation of the leukaemia-associated ETO homologous**.

3. RESULTS

In study that was performed by (Sofia Rondin et al, 2006) found that Acute myeloid leukaemia (AML) is commonly associated with balanced chromosomal translocations that fuse two unrelated genes. This results in the expression of an aberrant fusion protein. t(8;21) is one of the most common translocations found in patients with AML. It results in the expression of the chimaeric protein AML1-ETO. AML1 is a transcription factor of crucial importance during haematopoiesis. The function of the fusion partner eight-twenty-one (ETO) is much less understood. (Sofia Rondin et al, 2006) studied the interaction patterns of the ETO homologues as well as their expression pattern in haematopoietic cells. And she also examined the consequences of upregulation or downregulation of the proteins. She found that all the ETO homologues as well as AML1-ETO can interact with each other. She have also found that the ETO homologues, but not AML1-ETO can bind to the corepressor SIN3B. The proposed interactions of the ETO homologues might have implications for the onset of leukaemia, since it opens up for an AML1-ETO mediated disturbance of ETO homologue function as well as a regulation of AML1-ETO function by the ETO homologues. Examination of the expression patterns of ETO homologues in haematopoietic cells showed that the expression of ETO was restricted to erythroid cells, suggesting a role for ETO in erythropoiesis. MTG16 and MTGR1 are ubiquitously expressed in haematopoietic cells. The expression of MTG16 decreases during erythroid and granulocytic differentiation, suggesting a role for MTG16 in early haematopoiesis. The differential expression of the ETO homologues in haematopoietic cells implies a specific function for each protein in haematopoiesis. Attempts to knock-down MTG16 showed a discrepancy between RNA levels and protein levels, which could propose a mechanism to keep the expression of MTG16 constant.

Expression of the AML 1 - MTG8 fusion gene in t(8;21) AML study by (Hiroyuki Miyoshi et al, 1993) showed that the expression of the AML1, MTG8 and AML1-MTG8 genes was examined by Northern blot analysis of RNA isolated from blood tests from the t(8;21) AML patients, other AML patients and ordinary people, and a few hematopoietic cell lines. AnAML1-particular test (C6E3SS6) recognized four noteworthy transcripts in all examples inspected with the exception of the Raji cell line, albeit irregular bands were not plainly saw in the t(8;21) AML patients and the Kasumi-I cell line in view of various transcripts communicated at abnormal states (Figure 5). The declaration of AML1 is by all accounts constitutive at different phases of hematopoietic differentiation, since it was recognized in all hematopoietic cell lines inspected, particularly in myeloid ancestries (Figure 5 and unpublished information). Then again, utilizing a MTG8-particular segment of the AML1-MTG8 combination cDNA (CHI5H2S) as a test, two noteworthy transcripts of 6.2 and 7.8 kb were identified in Kasumi-I and every one of the four t(8;21) AML tests analyzed, yet not in typical fringe blood tests, a few cell lines without t(8;21) and the t(8;21) AML test abating. A 5.7 kb transcript was distinguished just in the Raji and HEL cell lines, conceivably relating to an ordinary MTG8 transcript.

Expression of the ETO homologues ETO is ubiquitously expressed in human tissues, albeit with a low expression in many of them. It is highly expressed in heart, brain and fetal brain (Wolford JK, et al, 1998). No ETO RNA is detected in peripheral blood (68). In haematopoietic cell lines, expression of ETO is restricted to the erythroleukaemic cell line HEL, myeloma cell lines and a few B-cell lines. In human bone marrow cells ETO is exclusively expressed in the erythroid lineage (Lindberg SR, et al, 2005). Both MTGR1 and MTG16 are ubiquitously expressed in human tissues. Except for the absence of MTG16 in kidney, both proteins are expressed in all tissues examined (Morohoshi F, et al, 2000). Also in most haematopoietic cell lines as well as bone marrow cells, both proteins are ubiquitously expressed, as opposed to the restricted expression of ETO in haematopoietic cells. Protein interactions of the ETO homologues, In 1998, three independent groups reported that ETO could bind to the corepressors N-CoR, SMRT and SIN3 and bind to histone deacetylases (HDACs) through the NHR4 domain (Wang J, et al, 1998). Recruitment of HDACs to a promoter closes the chromatin and makes it inaccessible for transcription. After these pioneering findings, it was established that several regions of ETO could bind to corepressors and different HDACs. In particular the NHR2 domain and the regions flanking it are important for binding of SIN3A (Amann JM, et al, 2001). ETO can bind to HDACs 1,2 and 3. The binding can either be direct or indirect through binding of SIN3 or N-CoR. The NHR2 important for corepressor interactions was also shown to be important for homo- or heterooligomerisation between the ETO homologues as well as AML1-ETO (Kitabayashi I, et al, 1998).

4. CONCLUSION

ETO homologue genes are commonly involved in reciprocal chromosomal translocation (t) characteristic of acute leukemia. For example, the ETO gene becomes fused to the AML1 (Runx1) transcription factor gene by t(8;21) resulting in the biosynthesis of the AML1-ETO fusion protein. Similarly, the MTG16 gene becomes fused to the AML1 gene by t(16;21) resulting in the production of the AML1-MTG16 fusion protein. The oncogenic fusion proteins interfere with hematopoietic gene regulation by transcriptional repression mediated by ETO and MTG16, respectively. Co-repressors-HDAC recruited by the ETO portion of AML1-ETO diminishes chromatin accessibility leading to transcriptional repression at AML1 targets, contributing to the cellular differentiation block of the leukemic cells.

REFERENCES

- [1] Gahrton, G. and B. Lundh, Blodsjukdomar. Lärobok i hematologi. 3rd ed. 1999, Stockholm: NATUR OCH KULTUR.
- [2] Mitelman, F., B. Johansson, and F. Mertens, Mitelman Database of Chromosome Aberrations in Cancer. 2006.
- [3] Rowley, J.D., Recurring chromosome abnormalities in leukemia and lymphoma. *Semin Hematol*, 1990. 27(2): p. 122-36.
- [4] Kelly, L.M. and D.G. Gilliland, Genetics of myeloid leukemias. *Annu Rev Genomics Hum Genet*, 2002. 3: p. 179-98.
- [5] Miyoshi H, Kozu T, Shimizu K, Enomoto K, Maseki N, Kaneko Y, et al. The t(8;21) translocation in acute myeloid leukemia results in production of an AML1-MTG8 fusion transcript. *Embo J* 1993;12(7):2715-21.
- [6] Zhang Y, Strissel P, Strick R, Chen J, Nucifora G, Le Beau MM, et al. Genomic DNA breakpoints in AML1/RUNX1 and ETO cluster with topoisomerase II DNA cleavage and DNase I hypersensitive sites in t(8;21) leukemia. *Proc Natl Acad Sci U S A* 2002;99(5):30705.
- [7] Levanon D, Negreanu V, Bernstein Y, Bar-Am I, Avivi L, Groner Y. AML1, AML2, and AML3, the human members of the runt domain gene-family: cDNA structure, expression, and chromosomal localization. *Genomics* 1994;23(2):425-32.
- [8] Look AT. Oncogenic transcription factors in the human acute leukemias. *Science* 1997;278(5340):1059-64.
- [9] Dash A, Gilliland DG. Molecular genetics of acute myeloid leukaemia. *Best Pract Res Clin Haematol* 2001;14(1):49-64.
- [10] Aplan PD. Causes of oncogenic chromosomal translocation. *Trends Genet* 2006;22(1):46-55.

- [11] Hagemeijer A, Garson OM, Kondo K. Fourth International Workshop on Chromosomes in Leukemia 1982: Translocation (8;21)(q22;q22) in acute nonlymphocytic leukemia. *Cancer Genet Cytogenet* 1984;11(3):284-7.
- [12] Dzierzak E, Medvinsky A. Mouse embryonic hematopoiesis. *Trends Genet.* 1995;11:359–366.
- [13] Gelmetti V, Zhang J, Fanelli M, Minucci S, Pelicci PG, Lazar MA. Aberrant recruitment of the nuclear receptor corepressor-histone deacetylase complex by the acute myeloid leukemia fusion partner ETO. *Molecular and cellular biology.* 1998;18(12):7185–7191.
- [14] Lutterbach B, Westendorf JJ, Linggi B, Patten A, Moniwa M, Davie JR, Huynh KD, Bardwell VJ, Lavinsky RM, Rosenfeld MG. ETO, a target of t(8;21) in acute leukemia, interacts with the N-CoR and mSin3 corepressors. *Molecular and cellular biology.* 1998;18(12):7176–7184.
- [15] Wang J, Hoshino T, Redner RL, Kajigaya S, Liu JM. ETO, fusion partner in t(8;21) acute myeloid leukemia, represses transcription by interaction with the human N-CoR/mSin3/HDAC1 complex. *Proceedings of the National Academy of Sciences of the United States of America.* 1998;95(18):10860–10865. doi: 10.1073/pnas.95.18.10860.
- [16] Hildebrand D, Tiefenbach J, Heinzel T, Grez M, Maurer AB. Multiple regions of ETO cooperate in transcriptional repression. *The Journal of biological chemistry.* 2001;276(13):9889–9895. doi: 10.1074/jbc.M010582200.
- [17] Amann JM, Nip J, Strom DK, Lutterbach B, Harada H, Lenny N, Downing JR, Meyers S, Hiebert SW. ETO, a target of t(8;21) in acute leukemia, makes distinct contacts with multiple histone deacetylases and binds mSin3A through its oligomerization domain. *Molecular and cellular biology.* 2001;21(19):6470–6483. doi: 10.1128/MCB.21.19.6470-6483.2001.
- [18] Zhang J, Hug BA, Huang EY, Chen CW, Gelmetti V, Maccarana M, Minucci S, Pelicci PG, Lazar MA. Oligomerization of ETO is obligatory for corepressor interaction. *Molecular and cellular biology.* 2001;21(1):156–163. doi: 10.1128/MCB.21.1.156-163.2001.
- [19] Erickson P, Gao J, Chang KS, Look T, Whisenant E, Raimondi S, Lasher R, Trujillo J, Rowley J, Drabkin H. Identification of breakpoints in t(8;21) acute myelogenous leukemia and isolation of a fusion transcript, AML1/ETO, with similarity to *Drosophila* segmentation gene, runt. *Blood.* 1992;80(7):1825–1831.
- [20] Miyoshi H, Kozu T, Shimizu K, Enomoto K, Maseki N, Kaneko Y, Kamada N, Ohki M. The t(8;21) translocation in acute myeloid leukemia results in production of an AML1-MTG8 fusion transcript. *The EMBO journal.* 1993;12(7):2715–2721.
- [21] Gamou T, Kitamura E, Hosoda F, Shimizu K, Shinohara K, Hayashi Y, Nagase T, Yokoyama Y, Ohki M. The partner gene of AML1 in t(16;21) myeloid malignancies is a novel member of the MTG8(ETO) family. *Blood.* 1998;91(11):4028–4037.
- [22] Wolford JK, Prochazka M. Structure and expression of the human MTG8/ETO gene. *Gene* 1998;212(1):103-9.
- [23] Lindberg SR, Olsson A, Persson AM, Olsson I. The Leukemia-associated ETO homologues are differently expressed during hematopoietic differentiation. *Exp Hematol* 2005;33(2):189-98.
- [24] Morohoshi F, Mitani S, Mitsuhashi N, Kitabayashi I, Takahashi E, Suzuki M, et al. Structure and expression pattern of a human MTG8/ETO family gene, MTGR1. *Gene* 2000;241(2):287-95.
- [25] Wang J, Hoshino T, Redner RL, Kajigaya S, Liu JM. ETO, fusion partner in t(8;21) acute myeloid leukemia, represses transcription by interaction with the human N-CoR/mSin3/HDAC1 complex. *Proc Natl Acad Sci U S A* 1998;95(18):10860-5.
- [26] Amann JM, Nip J, Strom DK, Lutterbach B, Harada H, Lenny N, et al. ETO, a target of t(8;21) in acute leukemia, makes distinct contacts with multiple histone deacetylases and binds mSin3A through its oligomerization domain. *Mol Cell Biol* 2001;21(19):6470-83.
- [27] Kitabayashi I, Ida K, Morohoshi F, Yokoyama A, Mitsuhashi N, Shimizu K, et al. The AML1-MTG8 leukemic fusion protein forms a complex with a novel member of the MTG8(ETO/CDR) family, MTGR1. *Mol Cell Biol* 1998;18(2):846-58.